

Relationships between silicon content and glutathione peroxidase activity in tissues of rats receiving lithium in drinking water

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Abstract Lithium salts are widely used in psychiatry, but their presence in organism can result in both beneficial and adverse effects. Silicon, the third most abundant trace element in humans as well as antioxidant enzyme glutathione peroxidase (GPx) play important roles in organism. The disturbance of their level can cause severe disorders. The aim of our work was to evaluate the influence of Li_2CO_3 administration in drinking water for a period of 4 weeks on Si content and GPx activity in the tissues of liver, kidney, brain and femoral muscle in rats. The concentrations of provided solutions were 0.7, 1.4, 2.6, 3.6, 7.1 and 10.7 mmol $\text{Li}^+ \cdot \text{dm}^{-3}$. GPx activity was decreased versus control as a consequence of Li treatment, particularly in kidney and brain. This effect could be suggested to contribute to renal abnormalities which could occur during Li therapy. Si tissue level was significantly enhanced versus control in liver and femoral muscle in groups receiving high Li doses. In brain no well-marked changes were observed, whereas in kidney we observed the depletion in low-Li-groups, restoration of Si level in higher-Li-groups and unexpected decrease in the highest-Li-group.

Positive correlations between Si content and GPx activity in the tissues of kidney ($r = 0.677$) and brain ($r = 0.790$) as well as negative correlation ($r = -0.819$) in femoral muscle were found. We consider that our results give some reason for suggesting that monitoring of silicon level in patients undergoing Li therapy could be recommended. However, more investigations should be performed, particularly regarding the relationships between Si and GPx in blood and urine Si excretion during lithium administration.

Keywords Silicon · Lithium · Glutathione peroxidase · Male rats

Abbreviations

GPx Glutathione peroxidase
ROS Reactive oxygen species

Introduction

Lithium is an element which, not being included into the essential bioelements, can influence numerous metabolic processes. It affects the activity of antioxidant enzymes (de Vasconcellos et al. 2006), “arachidonic acid cascade” enzymes (Bosetti et al. 2002) as well as phosphatases (Yenush et al. 2000). Lithium ion influences serotonergic (Subhash et al. 1999), dopaminergic

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(Al Banchaabouchi et al. 2004) and glutamate (Shaldubina et al. 2001) neuronal transmission. Li administration can affect functions of thyroid and parathyroid glands causing hypothyroidism (Engin et al. 2005) and hyperparathyroidism (Livingstone and Rampes 2006). Mutual relationships between lithium and both macro- and micro-elements homeostasis have already been reported (Carney and Jackson 1998; Tandon et al. 1999; Sharma and Iqbal 2005). Studies concerning different cytokines have displayed its complicated effect, resulting from the increase in both proinflammatory and negative immunoregulatory ones production (Maes et al. 1999; Szuster-Ciesielska et al. 2003). For almost 60 years lithium salts have been widely used in different fields of medicine. Their beneficial properties observed in cases of psychiatric disorders are known best of all (Fawcett 2003), however, recent years have brought reports revealing new possibilities of Li therapy application. Suppression of thyroid hormones caused by lithium made scientists study the efficacy of Li treatment as an adjuvant in patients suffering from thyroid diseases, undergoing radioiodine therapy (Vannucchi et al. 2005). Other authors have found the positive action of lithium in neurological disorders (Pardo et al. 2003). Topic application of lithium compounds have also been reported to show effectiveness in dermatological illnesses (Dreno et al. 2003). However, its presence in organism can also result in plenty of adverse effects, including renal and cardiovascular abnormalities, disturbances of cholesterol and glucose metabolism as well as abnormalities of erythropoiesis and leucopoiesis (Chmielnicka and Nasiadek 2003; Sharma and Iqbal 2005). It should be underlined that despite numerous works aiming at clearing of lithium's mechanism of action this question still remains unclear (Pardo et al. 2003).

Silicon is not so known as the other microelements however, after iron and zinc, it is the third most abundant trace element in human organism (Sripanyakorn et al. 2005). Si affects mainly the forming of connective tissues (Fauteux et al. 2005) but its presence has also been noted in blood and different organs (Bissé et al. 2005). It has been suggested to be requisite for lymphocyte proliferation in mammals (Seaborn et al. 2002).

Relationships between renal disturbances and silica exposure have been described (Governa et al. 2005).

Reactive oxygen species (ROS) are the products of uncompleted reduction of O_2 molecule (Drewa et al. 2002). They are formed in organisms during metabolic processes and can cause damage of proteins, membranes and DNA resulting in severe disturbances (Kuzniar et al. 2003; de Vasconcellos et al. 2006). ROS generation has been found to be involved into pathogenesis of the most dangerous diseases including cardiovascular abnormalities, Down syndrome and epilepsy (Gaeta et al. 2002; Kozlovski et al. 2006). Organisms have developed a complex defence system containing of high- and low-molecule substances which counteract ROS activity (Skibska et al. 2006). One of the most important antioxidant substances is glutathione peroxidase (GPx), an enzyme catalyzing the conversion of hydrogen peroxide into water. The correct action of antioxidant barrier is a question of great importance and depends on plenty of different factors. Alterations of antioxidants' level caused by abnormal elementary content of diet (Kuzniar et al. 2003) or by administration of different elements both under physiological and pathological conditions have been observed (Najda et al. 1994; Panneerselvam and Govindasamy 2004).

Having regarded the presented facts and aiming at contributing to knowledge of lithium's mechanism of action we tried to evaluate the influence of Li administration in drinking water on Si content and GPx activity in chosen tissues of rats.

Materials and methods

Our study was performed on 2-month-old male Wistar rats (180–220 g). The animals were divided into seven groups (six animals each): control group (C) received redistilled water, while six tested groups were given water solutions of Li_2CO_3 as the only drinking fluids. The concentrations were established as follows: 0.7, 1.4, 2.6, 3.6, 7.1 and 10.7 mmol $Li^+ \cdot dm^{-3}$. The animals had free access to standard feed LSM and drinking fluids. After four weeks rats were sacrificed under

ketamine narcosis. The tissues of liver, kidney, brain and femoral muscle were collected. 10% (w/v) tissue homogenates were prepared in Tris–HCl buffer ($0.1 \text{ mol} \cdot \text{dm}^{-3}$, $\text{pH} = 7.4$). Supernatants were obtained by centrifugation at $5000 \times g$ for 30 min. GPx activity was determined using RANSEL kit produced by RANDOX, according to Paglia and Valentine method (Paglia and Valentine 1967) and expressed in $\text{U} \cdot \text{g}^{-1}$ of protein. Protein was measured using method of Bradford (Bradford 1976). The concentration of silicon was measured using the spectrophotometric method (Wielkoszyński 2000). The assays were carried out with the help of SPECORD M40 (Zeiss Jena) spectrophotometer. Comparisons between control and tested groups were made using c-Cochran-Cox test. Values were considered significant with $P < 0.05$. The correlations between Si concentration and GPx activity were estimated with the help of the Pearson test.

The study was approved by I Local Ethical Commission of Feliks Skubiszewski Medical University of Lublin, acceptance 435/2003.

Results

The outcomes of the present experiment (Table 1) indicate that the influence of lithium administration on Si content and GPx activity depends strongly on the applied dose as well as on the tissue.

In liver and femoral muscle lower Li doses did not significantly alter silicon content. Higher Li

doses markedly increased Si versus control in both tissues. GPx activity was not significantly changed in both tissues versus control, except for liver in the group receiving the highest Li dose, where the statistically significant depletion was observed.

In brain Si content did not display well-marked changes versus control in all tested groups. GPx activity was decreased in dose-dependent way. In the group provided with the lowest dose the depletion was not statistically significant, while in the others the decrease showed statistical significance versus control.

The results concerning silicon content in kidney were unexpected. The administration of lower doses caused well-marked depression versus control. Higher ones caused the restoration of Si level to values observed in control group, whereas in the group receiving the highest dose of lithium the significant decrease versus control was obtained. GPx activity was markedly depressed versus control, without regard to the used dose.

The analysis of correlations between Si content and GPx activity displayed the existence of positive correlations in the tissues of kidney ($r = 0.677$) (Fig. 1) and brain ($r = 0.790$) (Fig. 2) as well as negative correlation ($r = -0.819$) in femoral muscle (Fig. 3).

Discussion

The present experiment revealed the deleterious impact of lithium administration on GPx activity,

Table 1 Silicon concentration and glutathione peroxidase activity in tissues of rats receiving lithium in drinking water

Li concentration (mmol dm^{-3})	Liver		Kidney		Brain		Femoral muscle	
	Si $\mu\text{mol g}^{-1}$ of fresh tissue $\bar{x} \pm \text{SD}$	GPx U g^{-1} of protein $\bar{x} \pm \text{SD}$	Si $\mu\text{mol g}^{-1}$ of fresh tissue $\bar{x} \pm \text{SD}$	GPx U g^{-1} of protein $\bar{x} \pm \text{SD}$	Si $\mu\text{mol g}^{-1}$ of fresh tissue $\bar{x} \pm \text{SD}$	GPx U g^{-1} of protein $\bar{x} \pm \text{SD}$	Si $\mu\text{mol g}^{-1}$ of fresh tissue $\bar{x} \pm \text{SD}$	GPx U g^{-1} of protein $\bar{x} \pm \text{SD}$
C	2.2 ± 0.5	1500 ± 424	3.5 ± 0.3	1300 ± 205	1.6 ± 0.4	172 ± 38	1.5 ± 0.7	77 ± 21
0.7	1.8 ± 0.6	1250 ± 278	1.9 ± 0.2^a	800 ± 182^a	1.4 ± 0.3	157 ± 34	2.3 ± 0.9	72 ± 26
1.4	1.9 ± 0.6	1300 ± 302	1.9 ± 0.4^a	674 ± 175^a	1.0 ± 0.4	87 ± 30^a	2.2 ± 0.7	67 ± 19
2.6	1.8 ± 0.6	1041 ± 215	2.0 ± 0.3^a	700 ± 164^a	1.1 ± 0.4	98 ± 24^a	2.8 ± 0.6	63 ± 20
3.6	3.5 ± 0.5^a	1066 ± 273	3.1 ± 0.9	754 ± 180^a	1.1 ± 0.4	90 ± 28^a	2.9 ± 0.9^a	61 ± 25
7.1	3.3 ± 0.5^a	1090 ± 302	3.1 ± 1.1	750 ± 107^a	1.2 ± 0.6	97 ± 22^a	3.3 ± 0.6^a	61 ± 22
10.7	3.4 ± 0.7^a	924 ± 254^a	2.2 ± 0.2^a	800 ± 124^a	1.2 ± 0.6	72 ± 27^a	3.1 ± 0.8^a	59 ± 21

$\bar{x} \pm \text{SD}$, mean \pm standard deviation

^a Statistical significance $P < 0.05$

Fig. 1 Positive correlation between GPx activity and Si content in the tissue of kidney

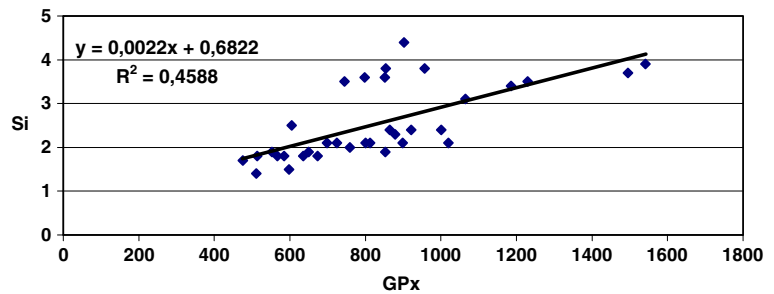


Fig. 2 Positive correlation between GPx activity and Si content in the tissue of brain

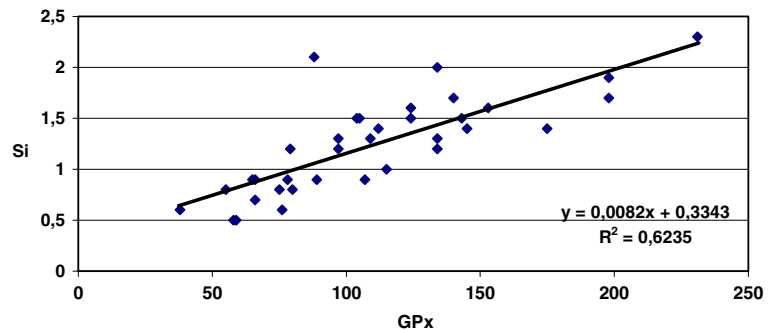
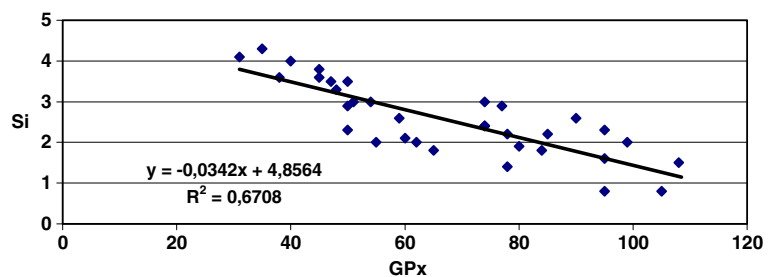


Fig. 3 Negative correlation between GPx activity and Si content in the tissue of femoral muscle



particularly in the tissues of kidney and brain. Our previous study concerned the influence of longer, 8-week administration of lithium on antioxidant enzymes. We noted that in that case GPx activity was insignificantly decreased in liver and brain, whereas in kidney well-marked depression versus control was obtained in all tested groups (Kiełczykowska et al. 2006b). The comparison of those results with the present ones points, that Li influence on GPx in brain is at least partially transient and can be overcome by organism, whereas in kidney the effect of Li exposure has more permanent character. The latter fact is consistent with the observed renal disturbances caused by lithium therapy (Chmielnicka and Nasiadek 2003; Livingstone and Rampes 2006). The outcomes obtained by other authors studying

Li influence on GPx have been proved to be inconsistent and the differences between action of lithium under physiological and pathological conditions have been found out.

It has been reported that increased ROS generation which occurs during immune system stimulation in phagocytic leukocytes can cause their damages. It encouraged the scientists to study the functions of antioxidant barrier in these cells under different conditions. LiCl given to sham-operated rats decreased GPx activity in neutrophils, whereas in bullectomised animals no significant effect was observed (Song et al. 1994).

Short-term administration of lithium carbonate to healthy rats resulted in no changes of GPx activity in erythrocytes and tissues of brain and liver (Abdalla and Bechara 1994).

Healthy rats provided orally with Li_2CO_3 for a period of 4 weeks showed a slight, insignificant GPx increase in liver, while in diabetic rats a very strong increase was displayed (Hu et al. 1999).

In rats undergoing LiCl + pilocarpine treatment inducing epilepticus status well-marked depletion of GPx in different regions of brain was reported (Eraković et al. 2000).

Lithium therapy is widely applied in patients suffering from different psychiatric diseases, exposed to stress. No significant influence of lithium on hippocampus GPx was observed in normal rats, whereas in animals submitted chronically to variate stress significant enhancement was shown (de Vasconcellos et al. 2006).

Kidney and liver were shown to belong to the main storage organs for silicon in rats (Almeida et al. 2002). Liver, kidney and brain functions could be influenced as a consequence of exposure to lithium (Chmielnicka and Nasiadek 2003). Relationships between Si and Li in pathological states were found (Bocca et al. 2005).

Lithium administration to rats resulted in divergent, dose-dependent influence on tissue silicon concentration. Our previous study, concerning the effect of longer, 8-week-exposure revealed different observations. The comparison of the outcomes point to the strong dependence between the time of exposure and the effect of lithium treatment. In liver in the first period of administration (4 weeks) higher doses of lithium caused significant increase, while after the longer exposure well-marked depression was obtained. In kidney after 8 weeks no significant influence was observed which suggests that in this organ lithium disturbs Si level transiently, at the beginning of treatment (Kiełczykowska et al. 2006a).

Our results provide some evidence for the existence of relationships between Si concentration and antioxidant defence in the chosen organs. The studies reveal links between oxidative processes and exogenous Si in plants (Liang et al. 2003) and animals (Kleczkowski et al. 2003). The addition of silicon alleviated oxidative stress in plants exposed to excess manganese (Shi et al. 2005). Exogenous silicon caused decrease of GPx activity in liver and kidney of rats (Najda et al. 1994). Antioxidants were found to have a preventive effect against silica-induced toxicity

(Zhang et al. 2000). However, there are not too many works concerning this question, so further investigations should be performed to provide its solution.

As a conclusion, it has been found that lithium administration has deleterious impact on GPx activity in important organs and this influence could contribute to adverse effects of Li therapy. Silicon homeostasis can also show abnormalities resulting from Li administration, although this question has not been studied systematically and further studies seem to be advisable, particularly in view of correlations between GPx activity and tissue Si content. We consider that our results give some reason for suggesting that monitoring of silicon level in patients undergoing Li therapy could be recommended. However, more investigations should be performed, particularly regarding the relationships between Si and GPx in blood and urine Si excretion during lithium administration.

References

- Abdalla DSP, Bechara EJH (1994) The effect of chlorpromazine and Li_2CO_3 on the superoxide dismutase and glutathione peroxidase activities of rat brain, liver and erythrocytes. *Biochem Mol Biol Int* 34:1085–1090
- Al Banchaabouchi M, Peña de Ortiz S, Menéndez R, Ren K, Maldonado-Vlaar CS (2004) Chronic lithium decreases Nurr1 expression in the rat brain and impairs spatial discrimination. *Pharmacol Biochem Behav* 79:607–621
- Almeida T, Soares ME, Cavalheiro J, de Lourdes Bastos M (2002) Silicon and iron levels in tissues of animals treated with a “ferrimagnetic ceramic” with oncotherapeutic potential (anti-tumor) value. *J Trace Elem Med Biol* 16:255–259
- Bissé E, Epting T, Beil A, Lindinger G, Lang H, Wieland H (2005) Reference values for serum silicon in adults. *Anal Biochem* 337:133–135
- Bocca B, Forte G, Petrucci F, Pino A, Marchione F, Bomboi G, Senofonte O, Giubilei F, Alimonti A (2005) Monitoring of chemical elements and oxidative damage in patients affected by Alzheimer’s disease. *Ann Ist Super Sanità* 41:197–203
- Bosetti F, Rintala J, Seemann R, Rosenberg TA, Contreras MA, Rapoport SI, Chang MC (2002) Chronic lithium downregulates cyclooxygenase-2 activity and prostaglandin E_2 concentration in rat brain. *Mol Psychiatry* 7:845–850
- Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein

- utilizing the principle of protein-dye binding. *Anal Biochem* 72:248–254
- Carney S, Jackson P (1998) Acute lithium administration impairs the action of parathyroid hormone on rat renal calcium, magnesium and phosphate transport. *Clin Exp Pharmacol Physiol* 25:795–799
- Chmielnicka J, Nasiadek M (2003) The trace elements in response to lithium intoxication in renal failure. *Ecotoxicol Environ Saf* 55:178–183
- de Vasconcellos APS, Nieto FB, Crema LM, Diehl LA, de Almeida LM, Prediger ME, da Rocha ER, Dalmaz C (2006) Chronic lithium treatment has antioxidant properties but does not prevent oxidative damage induced by chronic variate stress. *Neurochem Res* 31:1141–1151
- Dreno B, Chosidow O, Revuz J, Moyse D and the study investigator group. (2003) Lithium gluconate 8% vs. ketoconazole 2% in the treatment of seborrheic dermatitis: a multicentre, randomized study. *Br J Dermatol* 148:1230–1236
- Drewa G, Krzyżyńska-Malinowska E, Woźniak A, Protas-Drozd F, Mila-Kierzenkowska C, Rozwodowska M, Kowaliszyn B, Czajkowski R (2002) Activity of superoxide dismutase and catalase and the level of lipid peroxidation products reactive with TBA in patients with psoriasis. *Med Sci Monit* 8:BR338–BR343
- Engin A, Altan N, Isik E (2005) Erythrocyte glutathione levels in lithium-induced hypothyroidism. *Drugs R D* 6:35–40
- Eraković V, Župan G, Varljen J, Laginja J, Simonić A (2000) Lithium plus pilocarpine induced status epilepticus—biochemical changes. *Neurosci Res* 36:157–166
- Fauteux F, Rémus-Borel W, Menzies JG, Bélanger RR (2005) Silicon and plant disease resistance against pathogenic fungi. *FEMS Microbiol Lett* 249:1–6
- Fawcett JA (2003) Lithium combinations in acute and maintenance treatment of unipolar and bipolar depression. *J Clin Psychiatry* 64(Suppl 5):32–37
- Gaeta LM, Tozzi G, Pastore A, Federici G, Bertini E, Piemonte F (2002) Determination of superoxide dismutase and glutathione peroxidase activities in blood of healthy pediatric subjects. *Clin Chim Acta* 322:117–120
- Governa M, Amati M, Fenoglio I, Valentino M, Coloccini S, Bolognini L, Botta GC, Emanuelli M, Pierella F, Volpe AR, Astolfi P, Carmignani M, Fubini B (2005) Variability of biological effects of silicas: Different degrees of activation of the fifth component of complement by amorphous silicas. *Toxicol Appl Pharmacol* 208:68–77
- Hu M, Wu Y, Wu H (1999) Influence of streptozotocin-induced diabetes in rats on the lithium content of tissue and the effect of dietary lithium supplements on this diabetic condition. *Metabolism* 48:558–563
- Kielczykowska M, Musik I, Hordyjewska A, Lewandowska A, Pasternak K (2006a) The influence of different doses of lithium administered in drinking water on silicon concentration in plasma and chosen tissues of rats. *Pol J Environ Stud* 15:74–76
- Kielczykowska M, Pasternak K, Musik I, Wrońska-Tyra J, Hordyjewska A (2006b) The influence of different doses of lithium administered in drinking water on lipid peroxidation and the activity of antioxidant enzymes in rats. *Pol J Environ Stud* 15:747–751
- Kleczkowski M, Klucinski W, Sikora J, Zdanowicz M, Dziekan P (2003) Role of the antioxidants in the protection against oxidative stress in cattle—nonenzymatic mechanisms (Part 2). *Pol J Vet Sci* 6:301–308
- Kozlovski VI, Olszancki R, Chlopicki S (2006) Free radicals generated by xanthine/xanthine oxidase system augment nitric oxide synthase (NOS) and cyclooxygenase (COX)-independent component of bradykinin-induced vasodilatation in the isolated guinea pig heart. *Pharmacol Rep* 58:405–412
- Kuzniar A, Mitura P, Kurys P, Szymonik-Lesiuk S, Florianczyk B, Strycka-Zimmer M (2003) The influence of hypomagnesemia on erythrocyte antioxidant enzyme defence system in mice. *BioMetals* 16:349–357
- Liang Y, Chen Q, Liu Q, Zhang W, Ding R (2003) Exogenous silicon (Si) increases antioxidant enzyme activity and reduces lipid peroxidation in roots of salt-stressed barley (*Hordeum vulgare* L.). *J Plant Physiol* 160:1157–1164
- Livingstone C, Rampes H (2006) Lithium: a review of its metabolic adverse effects. *J Psychopharmacol* 20:347–355
- Maes M, Song C, Lin A-h, Pioli R, Kenis G, Kubera M, Bosmans E (1999) In vitro immunoregulatory effects of lithium in healthy volunteers. *Psychopharmacology* 143:401–407
- Najda J, Goss M, Gminski J, Weglarz L, Siemianowicz K, Olszowy Z (1994) The antioxidant enzymes activity in the conditions of systemic hypersilicemia. *Biol Trace Elem Res* 42:63–70
- Paglia DE, Valentine WN (1967) Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. *J Lab Clin Med* 70:158–168
- Panneerselvam SR, Govindasamy S (2004) Effect of sodium molybdate on the status of lipids, lipid peroxidation and antioxidant systems in alloxan-induced diabetic rats. *Clin Chim Acta* 345:93–98
- Pardo R, Andreolotti AG, Ramos B, Picatoste F, Claro E (2003) Opposed effects of lithium on the MEK-ERK pathway in neural cells: inhibition in astrocytes and stimulation in neurons by GSK3 independent mechanism. *J Neurochem* 87:417–426
- Seaborn CD, Briske-Anderson M, Nielsen FH (2002) An interaction between dietary silicon and arginine affects immune function indicated by con-A-induced DNA synthesis of rat splenic T-lymphocyte. *Biol Trace Elem Res* 87:133–142
- Shaldubina A, Agam G, Belmaker RH (2001) The mechanism of lithium action: state of the art, ten years later. *Prog Neuropsychopharmacol Biol Psychiatry* 25:855–866
- Sharma SD, Iqbal M (2005) Lithium induced toxicity in rats: a hematological, biochemical and histopathological study. *Biol Pharm Bull* 28:834–837
- Shi Q, Bao Z, Zhu Z, He Y, Qian Q, Yu J (2005) Silicon-mediated alleviation of Mn toxicity in *Cucumis sativus*

- in relation to activities of superoxide dismutase and ascorbate peroxidase. *Phytochemistry* 66:1551–1559
- Skibska B, Józefowicz-Okonkwo G, Gorąca A (2006) Protective effect of early administration of alpha-lipoic acid against lipopolysaccharide-induced plasma lipid peroxidation. *Pharmacol Rep* 58:399–404
- Song C, Killeen AA, Leonard BE (1994) Catalase, superoxide dismutase and glutathione peroxidase activity in neutrophils of sham-operated and olfactory-bulbectomised rats following chronic treatment with desipramine and lithium chloride. *Neuropsychobiology* 30:24–28
- Sripanyakorn S, Jugdaohsingh R, Thompson RPH, Powell JJ (2005) Dietary silicon and bone health. *Nutr Bull* 30:222–230
- Subhash MN, Vinod KY, Srinivas BN (1999) Differential effect of lithium on 5-HT₁ receptor-linked system in regions of rat brain. *Neurochem Int* 35:337–343
- Szuster-Ciesielska A, Tustanowska-Stachura A, Słotwińska M, Marmurowska-Michałowska H, Kandeferszerzeń M (2003) In vitro immunoregulatory effects of antidepressants in healthy volunteers. *Pol J Pharmacol* 55:353–362
- Tandon A, Nagpaul JP, Bandhu H, Singh N, Dhawan DK (1999) Effect of lithium on hepatic and serum elemental status under different dietary protein regimens. *Biol Trace Elem Res* 68:51–62
- Vannucchi G, Chiti A, Mannavola D, Dazzi D, Rodari M, Tadayyon S, Beck-Peccoz P, Fugazzola L (2005) Radioiodine treatment of non-toxic multinodular goitre: effects of combination with lithium. *Eur J Nucl Med Mol Imaging* 32:1081–1088
- Wielkoszyński T (2000) [Modified, spectrophotometric method of silicon determination in biological material]. *Diagn Lab* 36:377–385
- Yenush L, Bellés JM, López-Coronado JM, Gil-Mascarell R, Serrano R, Rodríguez PL (2000) A novel target of lithium therapy. *FEBS Lett* 467:321–325
- Zhang Z, Shen HM, Zhang QF, Ong CN (2000) Involvement of oxidative stress in crystalline silica-induced cytotoxicity and genotoxicity in rat alveolar macrophages. *Environ Res* 82:245–252